

# Nitrous oxide emissions in soils cropped with maize under long-term tillage and under permanent pasture in New Zealand

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## Abstract

Information on the influence of tillage on nitrous oxide (N<sub>2</sub>O) emissions is needed by researchers, policy makers and farmers for assessment of global ramifications of warming potential of greenhouse gas N<sub>2</sub>O. This research evaluates the effects of continuous long-term tillage and seasonal changes in N<sub>2</sub>O emissions in New Zealand temperate climate. The experiment was conducted in a Kairanga silty clay loam (Eutric Gleysol) where maize (*Zea mays* L.) was grown continuously for either 17 (K17) or 34 (K34) years, with conventional tillage. An adjacent permanent ryegrass (*Lolium perenne* L.) and clover (*Trifolium* sp.) pasture (PP) was used as a control. Mean N<sub>2</sub>O emissions under K17 and K34 and PP ranged from 0.04 to 1.35, 0.03 to 1.13, and 0.03 to 0.99 g N<sub>2</sub>O-N/ha/h, respectively. Average annual N<sub>2</sub>O emissions from K34 were slightly lower than those in the K17 fields although there were no differences in N<sub>2</sub>O emissions between PP and K17. Depleted organic C, total N and microbial biomass C in K34 as compared with K17, possibly limited denitrification process and decreased N<sub>2</sub>O emissions. Soil water content (SWC) was a key factor controlling N<sub>2</sub>O emissions in both pasture and cropped soils. Low emissions occurred when gravimetric SWC content was less than 30%. N<sub>2</sub>O emissions from grazed pasture were low during summer (dry) period and increased during winter (wet) season. There were no differences in N<sub>2</sub>O between the K17 and K34 during dry season but N<sub>2</sub>O emission rates in wet season were lower in the K34. Despite high temporal variability observed in the N<sub>2</sub>O emissions, our estimates based on low frequency of measurements over 1 year were in general agreement with those in the literature. © 2001 Elsevier Science B.V. All rights reserved.

**Keywords:** Greenhouse gas emissions; Soil organic carbon; Long-term conventional tillage; Seasonal changes; New Zealand

## 1. Introduction

There is some evidence that intensive tillage practices impact on the nitrogen (N) cycle through loss of

organic matter and deterioration of soil structure (Saggar et al., 2001; Shepherd et al., 2001), which could influence the extent of N<sub>2</sub>O emissions. It is known that soil disturbance through cultivation decreases N<sub>2</sub>O fluxes due to better aeration, while the amounts of fertiliser applied into cultivated soil increase the potential emissions of N<sub>2</sub>O (Shepherd, 1992). Mosier et al. (1997) found that changing

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cultivated soils back to grasslands eventually (after 8–50 years) led to N<sub>2</sub>O emissions similar to those of native soils of the same texture and parent material. They also found that conservation practices, aimed at reducing harmful effects of extensive conventional practices, contributed to N<sub>2</sub>O emissions through improved soil conditions. Increased soil organic matter, higher soil microbial biomass, and water content are conditions that favour emissions of N<sub>2</sub>O.

Colbourn (1988) observed denitrification in the undrained ploughed soil as one-fifth of that in drained direct-drilled soil. Tillage restricted denitrification to a greater extent than did drainage. The latter study attributed differences in denitrification to changes in soil structure caused by the tillage. The lack of disturbance in the reduced tillage soils leads to a reduction in large pores, increased soil aggregation and reduced soil aeration, therefore, produced higher denitrification.

Concentrations of biomass carbon (C) and organic C in the soil are highly correlated with denitrification activity (Drury et al., 1991), and there is evidence that increasing C availability decreases the N<sub>2</sub>O fraction (Firestone, 1982). Since organic matter in soils is represented by plant debris, crop species have a significant influence on the N<sub>2</sub>O emissions (Kaiser et al., 1998). The authors reported that the total N<sub>2</sub>O losses during the winter increased with the decreasing dry matter-to-N-content ratio of the plant residues incorporated into the soil by ploughing.

Ineson et al. (1998) demonstrated high levels of N<sub>2</sub>O production under elevated CO<sub>2</sub> conditions. Plants growing under elevated CO<sub>2</sub> may be subjected to greater N limitation because of the N being lost through denitrification. Ineson et al. (1998) recorded a 27% increase in N<sub>2</sub>O production under elevated CO<sub>2</sub> conditions, and attributed this increase to excess available soil C under elevated CO<sub>2</sub>. Plant roots may deplete available inorganic N and alter the soil physical status, resulting in decreased denitrification (Granli and Bockman, 1994). Denitrification rates generally increase with increasing soil water content (SWC). Sitaula and Bakken (1993) obtained a significant increase in N<sub>2</sub>O accumulation by increasing the SWC from 35 to 45%. Similarly, Carran et al. (1995) reported low or negligible N<sub>2</sub>O emissions in the Manawatu region, New Zealand, when the SWC was less than 35%.

Low N<sub>2</sub>O fluxes from grassland compared with cropped tillage systems also contribute to N<sub>2</sub>O

mitigation with additional benefits of restoration of soil properties (Mosier et al., 1997). On high-class soils in New Zealand (as classified by Webb et al., 1997 from well-defined, topographic, climatic and soil attributes), conversion of pasture for arable cropping is a common practice that generally results in deterioration of soil structure stability, and depletion of soil organic matter and N fertility over time (Saggar et al., 2001; Shepherd et al., 2001). Additional N fertilisers are then applied to compensate for the loss of N fertility and to maintain crop productivity. N fertilisers are considered a major source of N<sub>2</sub>O emissions.

In a previous study (Choudhary et al., 2001), we determined the rates of N<sub>2</sub>O emissions from plots established in 1995 sown with CT and no-tillage (NT), and with occasionally sheep-grazed permanent pasture (PP). We observed large inherent variations in N<sub>2</sub>O fluxes (a mean of coefficient of variation = 119%), probably reflecting natural soil heterogeneity. The N<sub>2</sub>O fluxes were substantially higher in CT and NT compared with those of PP, but no differences were found between CT and NT treatments. These plots did not have a long history of continuous tillage nor did the pasture plot represent a regularly grazed pasture common to New Zealand conditions.

Information on N<sub>2</sub>O emissions in medium- (17 years) and long-term (34 years) tilled soils may help to interpret the effects of cultivation on N-dynamics. The objective of this study was to determine the impacts of long-term continuous cropping with CT and seasonal changes on N<sub>2</sub>O emissions, and compare these with emissions from permanent pasture in temperate New Zealand and estimate annual average N<sub>2</sub>O emissions for these soils.

## 2. Materials and methods

### 2.1. Experimental site

The experimental site was at Kairanga (40°21'S, 175°39'E) located within the Manawatu region of the North Island of New Zealand. The soil was classified as Eutric Gleysol (FAO), Typic Endoaquept (US), Kairanga silt loam with very poor drainage and soil characteristics (Table 1) as described in Saggar et al. (2001). Three farms at this site representing a regularly fertilised grazed PP sown with introduced grasses

Table 1  
Land-use, symbol, soil classification, location, some soil characteristics of Kairanga silty clay loam soil used in this study

Site	Land use	Symbol	Soil classification			Grid reference <sup>a</sup> (NZMS 260)	pH 1:2.5 soil:water	Organic C (g/kg soil)	Total N (g/kg soil)	Microbial C (mg/kg soil)
			NZ	US	FAO					
Kairanga Silty clay loam <sup>c</sup>	Permanent	PP <sup>b</sup>	Typic	Typic	Eutric	S24 242 931	5.65 ± 0.0	49.9 ± 3.2	4.7 ± 0.9	1585 ± 5
	Pasture		Orthic Gley	Endoaquept	Gleysol		1 <sup>e</sup>			
	17 years	K17 <sup>d</sup>	Typic	Typic	Eutric	S24 229 948	5.58 ± 0.0	33.0 ± 4.9	2.8 ± 0.5	474 ± 11
	Maize		Orthic Gley	Endoaquept	Gleysol		4			
	34 years	K34 <sup>d</sup>	Typic	Typic	Eutric	S24 243 093	6.20 ± 0.0	20.3 ± 3.0	1.8 ± 0.9	261 ± 15
	Maize		Orthic Gley	Endoaquept	Gleysol		2			

<sup>a</sup> Specifies the farm easting and westing location on New Zealand Map Series (NZMS).

<sup>b</sup> Site renewed 8 years ago (1989) following a wheat (*Triticum* sp.); before this under long-term ryegrass (*Lolium perenne* L.) and clover (*Trifolium* sp.).

<sup>c</sup> Particle size distribution (g kg<sup>-1</sup>); Kairanga: sand 90–120, silt 510–530, clay 400–420.

<sup>d</sup> Sites K17 and K34 were under continuous maize (*Zea mays* L.).

<sup>e</sup> Values represent mean ± standard error.

(*L. perenne* L.) and legumes (*Trifolium* sp.); a 17-years conventional cultivation (K17), and a 34-years conventional cultivation (K34) grown maize (*Z. mays*) crop were included in this study. The PP was intensively grazed by dairy stock (160 cows strip-grazed on 1.21 ha sections of the farm for 9 h during the day and 12 h during the night), and was used as a control. Both cropping sites (K17 and K34) were conventionally tilled, up to the 250 mm soil depth using a mould-board plough followed by levelling and rolling, disking and power harrowing.

Maize was sown at both cultivated farms in late October 1998. The K34 farm received 306.5 kg/ha of (15:10:10:8) fertiliser at sowing and additional 300 kg/ha of urea as side-dressing applied in early December 1998. The maize crop harvested for silage produced 16 Mg DM/ha. The K17 farm received 153 kg/ha (half of K34) of (15:10:10:8) starter fertiliser applied at the end of October and 300 kg/ha of urea side-dressed in early December 1998. The yield of maize grain was 6.9 Mg/ha. Coupled with a dry summer (which followed a very wet October), the fertilisers (especially the urea side-dressing) were not utilised effectively, because of the lack of rainfall and very dry soils. Urea pellets were still visible 2 weeks after application.

## 2.2. Soil sampling

Within each site, four random locations were chosen, and 25 cores (25 mm dia, 0–10 and 10–20 cm depths) were taken at approximately even spacings in an area of ca 50 m<sup>2</sup> in September 1998, before spring cultivation and sowing. The cores were pooled, field moist soils (–10 kPa) were sieved (<2 mm) soon after collection, and sieved soils stored at 4°C were used for soil microbial biomass C analysis. Subsamples of the sieved soil were air dried and used for pH, C and N analyses.

## 2.3. Soil analyses

Soil microbial biomass C was determined by a fumigation–extraction method. Fumigated and non-fumigated soils were extracted with 0.5 M K<sub>2</sub>SO<sub>4</sub> for 30 min (1:5 soil:extractant ratio), and the extracts analysed for C by TOC 5000 (Shimadzu, Kyoto) analyser. The C obtained from the fumigated samples

minus that from the non-fumigated samples was taken to represent the microbial-C flush and converted to microbial-biomass C using the relationship: Microbial C=C flush/0.41.

Air-dried samples were used for the total C and N analyses. Total C in soils was analysed by a combustion method (Induction Furnace, Leco, St. Joseph, MI). Total soil N was determined by semi-micro Kjeldahl digestion followed by NH<sub>4</sub><sup>+</sup>-N measurements in the digests by an autoanalyser procedure. Soil pH (1:2.5 water) was determined according to Blakemore et al. (1987).

Soil water content and temperature were measured at 100 mm depth each time during N<sub>2</sub>O measurement. Soil samples were collected, weighed, oven-dried to constant mass at 105°C, and weighed again. The final mass  $M_s$ , and the difference between fresh and dry masses  $M_w$  were used to calculate the gravimetric SWC:

$$\text{SWC} = \left( \frac{M_w}{M_s} \right) \times 100 \quad (1)$$

Soil temperature was measured with a handheld digital thermometer. A temperature probe was inserted into the soil next to each chamber and temperature indications were recorded manually. Particle size distribution was determined by pipette method.

## 2.4. Nitrous oxide measurements

Monthly, and sometimes fortnightly, N<sub>2</sub>O emission measurements included the cropped (October–May) as well as the winter fallow period (June–September). Measurements started in November 1998 and continued until September 1999. During this period a total of 14 measurements were made, although one data set collected on March 31 could not be analysed due to instrument malfunction. Rainfall data for the 3-year period (1996–1999) is shown in Fig. 1. The sites have a temperate humid climate with mean annual air temperatures of 12.1°C. The mean monthly temperature ranged between 15 and 20°C during summer, and between 8.4 and 10.2°C during winter.

Nitrous oxide emissions were measured following the chamber technique developed by Mosier and Hutchinson (1981). Each chamber was inserted 100 mm into the soil. Headspace gas samples were collected using 60 ml polypropylene syringes fitted

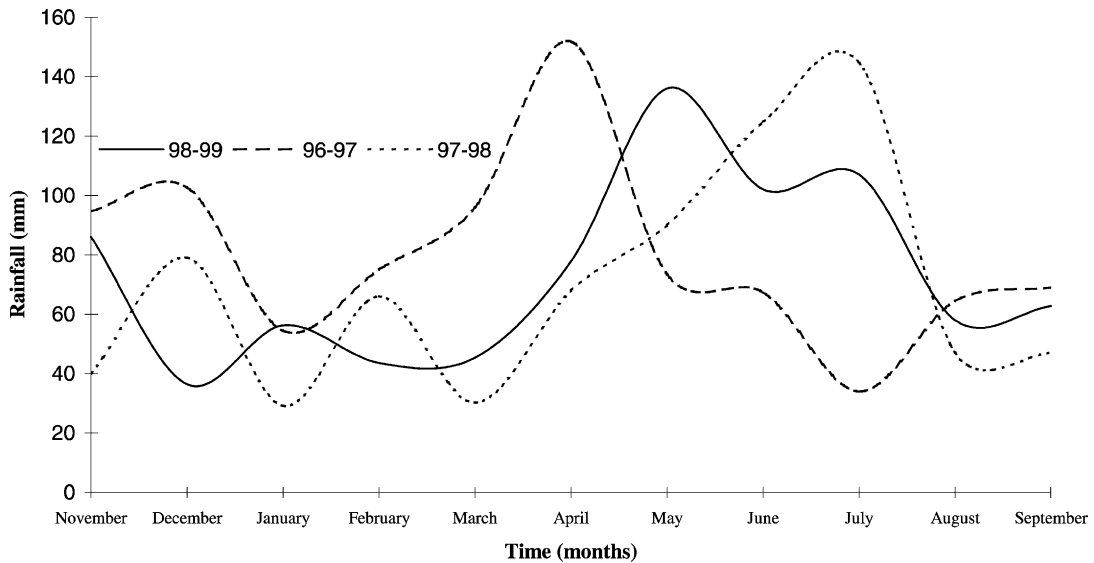


Fig. 1. Monthly rainfall from November 1998 to September 1999, 1996–1997, and 1997–1998.

with 3-way taps. Because of concern over the permeability of polypropylene to  $N_2O$ , field samples were analysed as soon as possible after collection, usually within 24 h. Initially, the chambers were left for 3 h with 1 h interval for gas sampling in order to detect any leakage. In later sampling, gas was collected only for 2 h. Analyses were done by gas chromatography (Mosier and Mack, 1980).

### 2.5. Statistical analysis

Unless otherwise stated, soil chemical and biochemical results were expressed on the basis of the oven-dry (105°C) weight of the material. A general linear model procedure (GLM) was used for analysis of experimental data. An analysis of variance (ANOVA) using a test of least significant difference (LSD) at 5% confidence level was used for comparison of treatments. Correlation of SWC and  $N_2O$  fluxes was analysed by mixed model procedure.

## 3. Results and discussion

### 3.1. Tillage effect on field $N_2O$ emissions

Initial analysis of data showed significant variability (data not shown), as discussed elsewhere (Choudhary

et al., 2001). Because the data were not normally distributed, comparison between treatments was accomplished by log-transformed data. The estimated annual  $N_2O$  emissions from the PP, K17 and K34 (calculated as the mean of all individual measurements of the closed cover chamber between November 1998 and September 1999) were similar at 3.24, 3.42 and 2.37 kg  $N_2O$ -N/ha/year, respectively (Table 2). Mean emission rates for the PP were within the range reported by Carran et al. (1995) from similar grazed sites on Kairanga soil. The  $N_2O$  emissions from the K17 were slightly higher than those from the K34 fields ( $P = 0.1$ ), and were comparable with those recorded by Jacinthe and Dick (1997) for a similar continuous maize system in Ohio, USA. Similar to our recent  $N_2O$  flux measurements under CT and NT (Choudhary et al., 2001), spatial variations in  $N_2O$  emissions were large. The mean flux rates ranged from 0.175 to 13.32, 0.175 to 16.91 and 0.088 to 30.05 kg  $N_2O$ -N/ha/year in PP, K17 and K34, respectively.

Although overall comparison of treatment means did not show any discernible differences between management practices, the trends in data suggested that the K34 had lower emissions ( $P = 0.1$ ) compared with those in the PP (Table 2). During early crop growth (November and December), the management practices had little effect on  $N_2O$  emissions (Fig. 2c). During the first 8 weeks of measurements,  $N_2O$  fluxes

Table 2  
The effects of tillage systems on the means of N<sub>2</sub>O emissions from the Kairanga field sites

Treatment	N <sub>2</sub> O emissions		
	Per hour (g N <sub>2</sub> O-N/ha)	Per day (g N <sub>2</sub> O-N/ha)	Per year (kg N <sub>2</sub> O-N/ha)
PP <sup>a</sup>	0.37	8.88	3.24
K17 <sup>b</sup>	0.39	9.36	3.42
K34 <sup>b</sup>	0.27	6.48	2.37
LSD <sub>0.05</sub>	ns <sup>c</sup>	ns	ns

<sup>a</sup> Permanent pasture.

<sup>b</sup> 17 and 34 years of continuous conventional grown maize, respectively.

<sup>c</sup> Not significant.

from the K17 treatment were lower than those in the K34 field, which probably resulted from lower application rate (153 kg/ha) of the starter N fertiliser to the K17 field at sowing. Additional side-dressing of 300 kg/ha of urea in early December did not appear to have had much effect on N<sub>2</sub>O emissions. Coupled with the very dry summer (which followed the wet October), the fertiliser added (especially the urea side-dressing) was probably not utilised effectively because of lack of rainfall and resulting dry soils (T.G. Shepherd, pers. commun., 2000). Some urea granules were still visible on the soil surface 2 weeks after application.

In January, the effects of management practices started to show when N<sub>2</sub>O emissions in the PP reached the lowest flux (approximately 0.1–0.2 mg N<sub>2</sub>O-N/ha/day), due to rapid drying of soil in the December–February summer period (data not shown). On the other hand, N<sub>2</sub>O fluxes remained high in the K17 and K34 fields (0.4–1.4 and 0.7–1.4 mg N<sub>2</sub>O-N/ha/day, respectively) indicating that during the maize growth period in summer, biological activities remained high.

In February, N<sub>2</sub>O emissions decreased in the K17. Similarly, in the K34 field, N<sub>2</sub>O fluxes started to drop sharply in March. Rainfall was also low in February and March (Fig. 1). However, the 8.5 mm rain before sampling on March 15, and the 9.5 mm that fell during the day, seemed to have created a burst of N<sub>2</sub>O flux from PP: It reached almost 2.4 mg N<sub>2</sub>O-N/ha/day, the highest emission rate recorded during the sampling period. The steady decrease of N<sub>2</sub>O emissions in the K17 and K34 fields coincided with crop maturation (Fig. 2c). The decrease was more pronounced in the K34 after harvest. Maize in the K34 field was harvested for silage on March 17 and 18. Maize in the

K17 resembled that in the K34 field, and was senescent during autumn.

In the cropped soils, N<sub>2</sub>O emissions decreased after March, although the PP maintained higher fluxes (Fig. 2c). More frequent rainfall events during April and May (Fig. 1) did not seem to have affected N<sub>2</sub>O emissions in the K17 and K34 fields, the emissions remained the lowest (Fig. 2c). There are several likely explanations for this phenomenon. It was probable that little precipitation led to low soil water, thus creating denitrification. Denitrification measured by using acetylene inhibition at the same sites indicated that denitrification rates were high (B.K. Daly, pers. commun., 2000). Alternatively, low soil nitrate levels, low water content in the soil, and low C input through root growth by the senescent crop perhaps slowed down biological activities in the soil, resulting in reduced N<sub>2</sub>O emissions.

Grain maize in the K17 was harvested on 19 May, with crop residue left on the soil surface. Also on 19 May sampling was conducted, following a rainfall event (30 mm on 17 May) 2 days before, but the expected flush of N<sub>2</sub>O fluxes did not occur (Fig. 2c). To determine if the N<sub>2</sub>O fluxes were affected after 4 days, another sampling was carried on 23 May. However, no such differences in nitrous gas emissions were found between the two sampling dates in K17 and K34 (Fig. 2c). This may be due to the unchanged soil conditions: although soil temperature in the K17 decreased moderately (Fig. 2b), SWC in K17 did not differ much due to the surface residue left in the field (Fig. 2a). However, a slight decrease in SWC in the PP led to 41% reduction in N<sub>2</sub>O fluxes. This indicates that SWC was a governing factor in N<sub>2</sub>O emissions in PP.

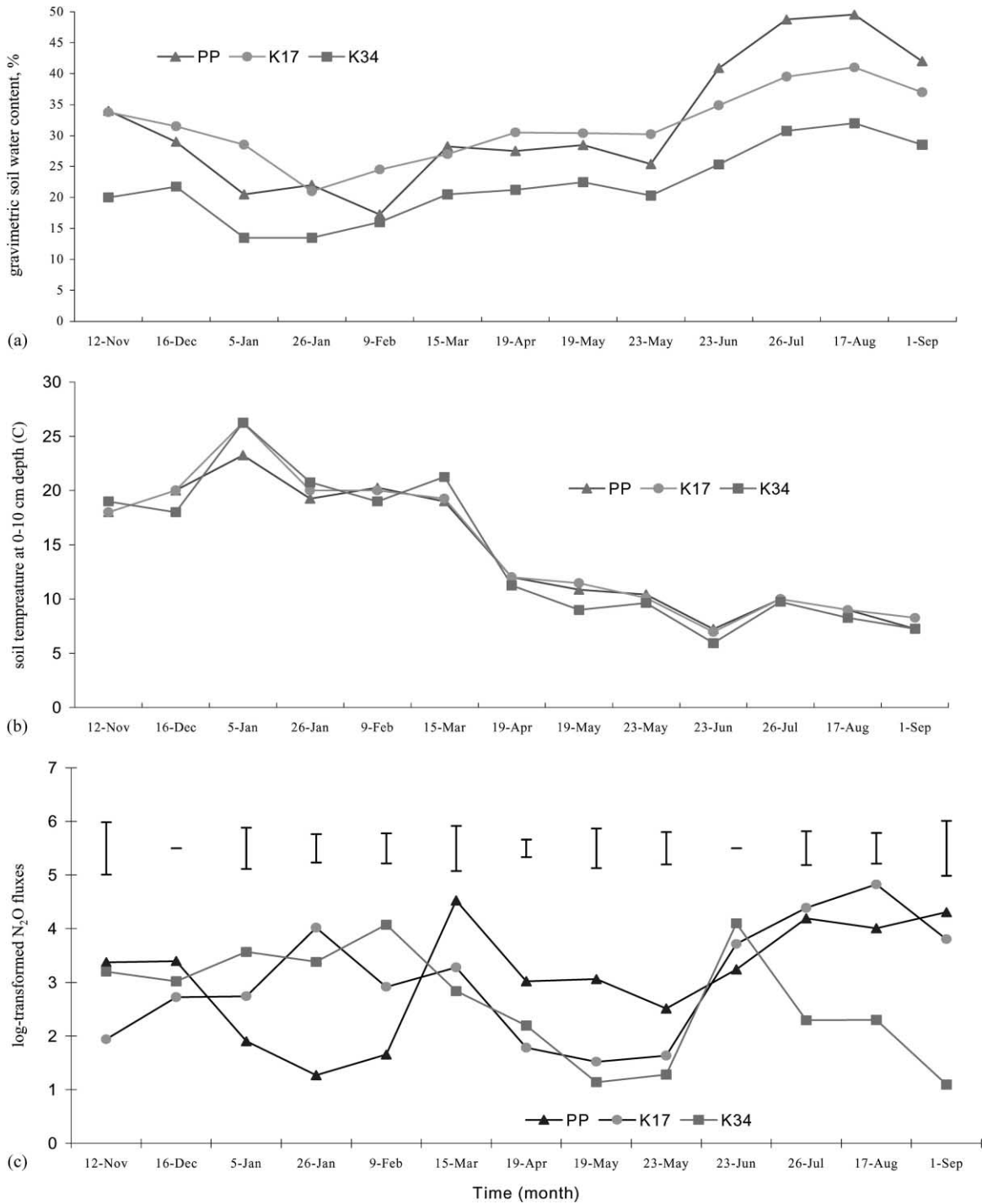


Fig. 2. The effect of PP, 17-year continuous cultivation (K17) and 34-year continuous cultivation (K34) on: (a) SWC; (b) soil temperature; and (c) log-transformed means of N<sub>2</sub>O emissions. (vertical bars represent LSDs).

During the fallow in June there was a sudden significant increase in  $N_2O$  emissions in both the K17 and K34 fields (Fig. 2c). A more steady increase was observed in the PP. With the onset of winter and a colder period, soil temperature dropped (Fig. 2b), and significant differences became apparent between the treatments.  $N_2O$  fluxes declined and returned to rates similar to those observed during autumn in the K34, whereas the PP and K17 field emissions steadily increased. The explanation for such behaviour of soil  $N_2O$  fluxes is rather complicated without measurements of additional parameters (such as soil ammonium and nitrate levels, microbial biomass, substrate induced respiration, etc.). Based on the limited available data, low emissions during winter were possibly due to low temperatures (Fig. 2b). On the other hand, interestingly, the highest fluxes measured were during winter, and were 2.7 and 2.2 mg  $N_2O$ -N/ha/day in the K34 and K17, respectively. These high  $N_2O$  fluxes probably reflected enhanced denitrification due to higher rainfall.

The higher organic C content plus surface residue in the K17 field (Table 1) could indicate that available C limits microbial  $N_2O$  production during winter (Kaiser et al., 1998). These authors observed a significant relationship between total  $N_2O$  emissions during winter and the ratio of dry matter-to-N content of the incorporated plant residues. The K34 had low soil organic matter content (Table 1) and, as the maize crop was harvested for silage, this left little surface residue input. When considering the  $N_2O$  emission rates after harvest in March, with the exception of the  $N_2O$  emissions burst in June, it is probable that low soil nitrate levels of applied fertiliser already utilised by the maize in K34, were the determining factors in governing emissions. Moreover, given the favourable SWC (Fig. 2a), the  $N_2O$  fluxes were still low despite other researchers reporting high emissions in winter (Kessavalou et al., 1998). This indicates that in this case soil nitrate level may be limiting. However, there is a need to establish the effect of other soil parameters on  $N_2O$  emissions.

The  $N_2O$  emissions measured from PP were higher than those reported for occasionally sheep-grazed pasture (Choudhary et al., 2001). These relatively high  $N_2O$  fluxes appeared to be associated with the pastoral management system used. The PP used in this study was intensively grazed by dairy stock, and could

have larger urine returns than the occasionally sheep-grazed pasture reported by Choudhary et al. (2001). Ruz-Jerez et al. (1994) and Williams et al. (1999) have already stressed the influence of grazing animals on input of readily decomposable excreta. Considering that urine is a concentrated N solution (approx. 10 g N/l, of which 80–90% is urea), and the effective rate of application within urine patches is often greater than the equivalent of 500 kg N/ha (Haynes and Sherlock, 1986), denitrification losses could be 30–40% higher (Carran et al., 1982). High N inputs in the form of urine were apparently beyond the ability of plant uptake.

### 3.2. Effect of SWC on $N_2O$ emissions

Gravimetric SWC and soil temperature measurements from the vicinity of the installed chambers are shown in Fig. 2a and b. The SWC was significantly affected by tillage treatments throughout the year, although there were no differences in soil temperature. The K34 field SWC was 22–52% lower than that in the K17 field. The pattern of SWC changes in K17 closely resembled the pattern in K34. The SWC in the PP was generally higher than that in the K34. There were no marked differences in SWC between the K17 and PP fields during the year except the winter period (June to early September). Winter SWC in the PP was substantially higher compared with the K17 and K34 fields.

The effects of SWC on  $N_2O$  emissions in the K17 and PP were different from those in the K34 (Fig. 3). In the K34 field, SMC did not seem to govern fluxes which were more apparent during wet months of April and May. The regression analyses using the measured data revealed no relationship between the SWC and  $N_2O$  fluxes in the K34 treatment. Nevertheless, the log-transformed values of  $N_2O$  emissions showed a low negative correlation ( $r = -0.42$ ) between the two parameters (Fig. 3). However,  $N_2O$  fluxes from PP and K17 appeared to be influenced by SWC ( $r = 0.72$  and  $0.26$ , respectively). There is clear indication (Fig. 3) that low emissions occur when gravimetric SWC content is less than 30%. Above this apparent threshold, the upper level of emission values increases with water content, but low values still occur frequently, as observed by Carran et al. (1995). Although  $N_2O$  fluxes did not follow the rainfall patterns, regression analysis



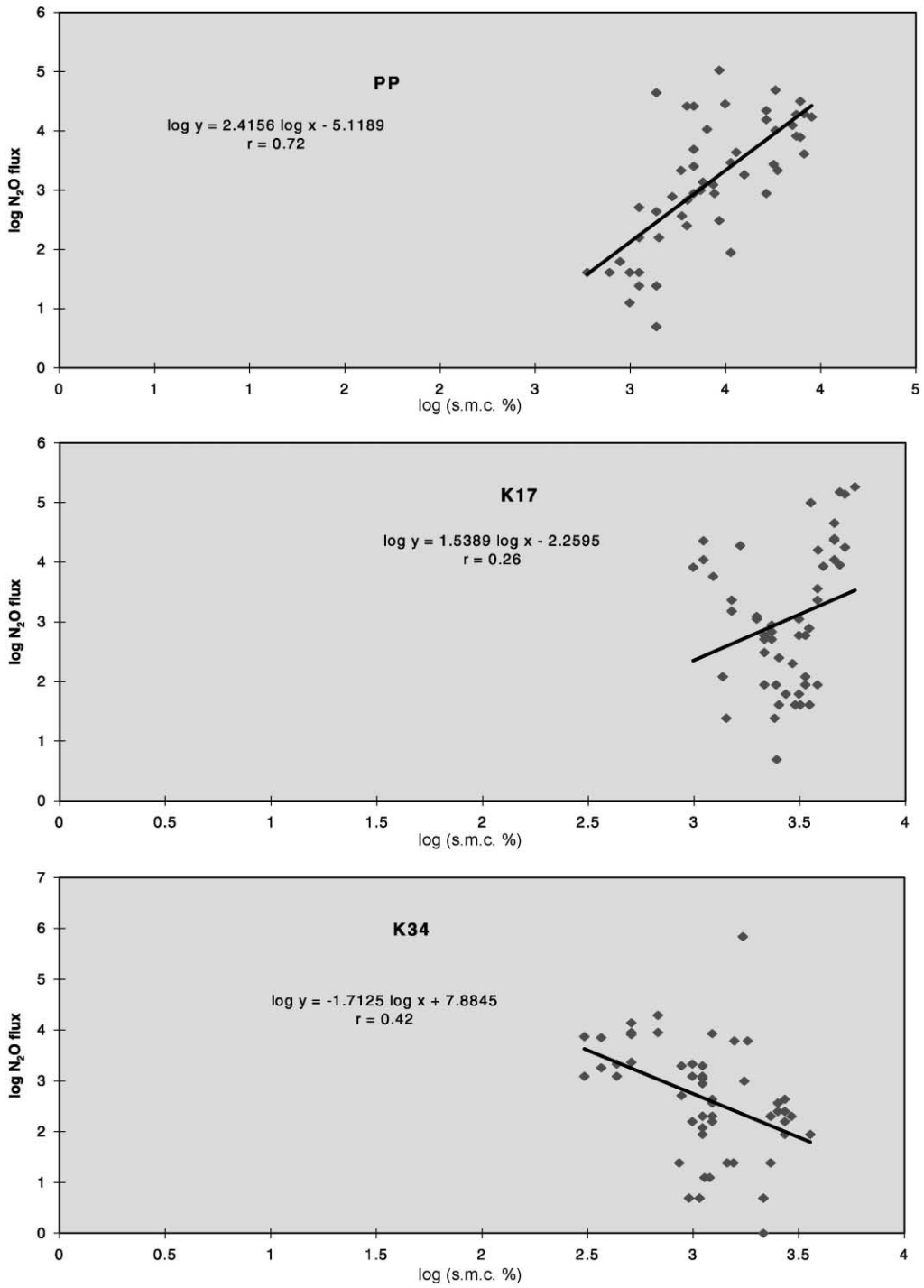


Fig. 3. Regression analysis of log-transformed data between SWC and N<sub>2</sub>O emissions in PP, 17-year continuous cultivation (K17) and 34-year continuous cultivation (K34) soils.

Table 3  
Seasonal log-transformed field N<sub>2</sub>O emissions in the Kairanga field sites<sup>a</sup>

Treatment	Spring (September–November)	Summer (December–January–February)	Autumn (March–April–May)	Winter (June–July–August)	LSD <sub>0.05</sub>
PP <sup>b</sup>	3.91 a	2.27 b	3.46 a	3.81 a	0.64
K17 <sup>c</sup>	3.01 b	3.08 b	2.23 c	4.31 a	0.66
K34 <sup>c</sup>	2.32 bc	3.53 a	2.09 c	2.90 ab	0.75

<sup>a</sup> Values followed by the same letter in rows show no significant differences ( $P < 0.05$ ).

<sup>b</sup> Permanent pasture.

<sup>c</sup> 17 and 34 years of continuous conventional grown maize, respectively.

using log-transformed data indicated a low but significant relationship with SWC. These results concur with those reported by Kaiser et al. (1998) for arable soils and with Carran et al. (1995) for pasture sites in these soils.

### 3.3. Effects of seasonal variations on N<sub>2</sub>O emissions

General seasonal patterns of N<sub>2</sub>O emissions were well defined for the PP, but less pronounced for the K17 and K34 (Table 3). The pattern of N<sub>2</sub>O emissions from the PP are consistent with the data of Carran et al. (1995) for the same soil type, and the values for the wet season are similar to those recorded by Ruz-Jerez et al. (1994) in low fertility soils.

Seasonal grouping of monthly log-transformed N<sub>2</sub>O emissions showed significant differences in all treatments (Table 3). Summer season N<sub>2</sub>O emissions in the PP were lower than those in other seasons, whereas no discernible differences were observed in other seasons. Although N<sub>2</sub>O fluxes during spring and summer were similar in the K17 field, they were significantly lower than the winter fluxes, and higher than the autumn ones. There were considerably higher emissions in summer than in autumn in the K34, but seasonal variation between winter and spring was less profound. Mixed N<sub>2</sub>O flux response to seasonal variations between treatments makes it difficult to interpret these data. Similar patterns in gas emissions in the K17 and PP during the wet season suggest that SWC is a key factor controlling N<sub>2</sub>O emissions in soils. Carran et al. (1995) postulated that the absence of significant emission events while the soil is dominantly aerobic suggest that nitrification or other transformations of urine-derived N do not contribute to overall emissions in an important way. This pattern

suggests that denitrification was the primary source of N<sub>2</sub>O in the PP.

The onset of wet season can be early, as in 1996–1997, or late as in 1997–1998 (Fig. 1). While wet season fluxes in the K34 were generally low, high rainfall certainly influenced N<sub>2</sub>O emissions in the K17 and PP fields. As suggested earlier, losses in N<sub>2</sub>O during the wet season may form a major portion of total annual emissions from grazed pasture and cropped fields.

## 4. Conclusions

Annual N<sub>2</sub>O fluxes ranged widely from 0.18 to 13.32, 0.18 to 16.91, and 0.09 to 30.05 kg N<sub>2</sub>O-N/ha in the PP, K17 and K34 fields, respectively. Overall average annual N<sub>2</sub>O emissions from the K34 (2.37 kg N<sub>2</sub>O-N/ha) were lower at ( $P = 0.1$ ) than the K17 fields (3.42 kg N<sub>2</sub>O-N/ha). The latter were similar to emissions from pasture (3.24 kg N<sub>2</sub>O-N/ha). Depleted total C and N content in the K34, which were low due to continuous intensive cropping, low surface residue (as DM removed for silage), and decreased water holding capacity, as compared with K17, could have limited the denitrification process.

In accordance with previous studies (Ruz-Jerez et al., 1994; Carran et al., 1995), we found N<sub>2</sub>O emissions from grazed pasture were low during the summer (dry) period and increased during the winter (wet) season. There were no differences in N<sub>2</sub>O between the K17 and K34 during the summer season but N<sub>2</sub>O emission rates in winter were lower in the K34.

This study represents the first New Zealand comparison of N<sub>2</sub>O emission estimates from long-term

cropped and permanent pasture soils. While the uncertainties associated with these estimates are likely to be large due to high temporal variability observed in the N<sub>2</sub>O emissions, our estimates based on low frequency of measurements (13 sampling) over 1 year were in general agreement with those in the literature. Bouwman (1996) showed that N<sub>2</sub>O emission estimates from agricultural soils increase as the frequency of measurement increases. Therefore, further prolonged studies with a high frequency of sampling are required to confirm and obtain a robust average yearly value for pasture and cropped soils in New Zealand.

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### References

- Blakemore, L.C., Searle, P.L., Daly, B.K., 1987. Methods for chemical analysis of soils. New Zealand Soil Bureau Scientific Report 80, 103 pp.
- Bouwman, A.F., 1996. Direct emission of nitrous oxide from agricultural soils. *Nutr. Cycl. Agroecosyst.* 46, 53–70.
- Carran, R.A., Ball, P.R., Theobald, P.W., Collins, M.E.G., 1982. Soil nitrogen balances in urine-affected areas two moisture regimes in Southland. *NZ J. Exp. Agric.* 10, 377–381.
- Carran, R.A., Theobald, P.W., Evans, J.P., 1995. Emission of nitrous oxide from some grazed pasture soils in New Zealand. *Aust. J. Soil Res.* 33, 341–352.
- Choudhary, M.A., Akramkhanov, A., Saggarr, S., 2001. Nitrous oxide emissions from cropped fields: tillage effects, spatial and seasonal variability. *Agric. Ecosyst. Environ.*, in revision.
- Colbourn, P., 1988. The influence of drainage and cultivation on denitrification losses from an arable clay soil. In: Jenkinson, D.S., Smith, K.A. (Eds.), *Nitrogen Efficiency in Agricultural Soils*. Elsevier, New York, pp. 283–294.
- Drury, C.F., McKenney, D.J., Findlay, W.I., 1991. Relationships between denitrification, microbial biomass and indigenous soil properties. *Soil Biol. Biochem.* 23, 751–755.
- Firestone, M.K., 1982. Biological denitrification. In: Stevenson, F.J. (Ed.), *Nitrogen in Agricultural Soils*. Agronomy Monograph No. 22. American Society of Agronomy, Madison, WI, pp. 289–326.
- Granli, T., Bockman, O.C., 1994. Nitrous oxide from agriculture. *Norwegian J. Agric. Sci. (Suppl.)* 12, 7–128.
- Haynes, R.J., Sherlock, R.R., 1986. Gaseous losses of nitrogen. In: Haynes, R.J. (Ed.), *Mineral Nitrogen in the Plant–Soil System*. Academic Press, New York, pp. 242–302.
- Ineson, P., Coward, P.A., Hartwig, U.A., 1998. Soil gas fluxes of N<sub>2</sub>O, CH<sub>4</sub> and CO<sub>2</sub> beneath *Lolium perenne* under elevated CO<sub>2</sub>: the Swiss free air CO<sub>2</sub> enrichment experiment. *Plant Soil* 198, 89–95.
- Jacinthe, P.A., Dick, W.A., 1997. Soil management and nitrous oxide emissions from cultivated fields in southern Ohio. *Soil Till. Res.* 41, 221–235.
- Kaiser, E.A., Kohrs, K., Kucke, M., Schnug, E., Heinemeyer, O., Munch, J.C., 1998. Nitrous oxide release from arable soil: importance of N-fertilisation, crops and temporal variation. *Soil Biol. Biochem.* 30, 1553–1563.
- Kessavalou, A., Doran, J.W., Mosier, A.R., Drijber, R.A., 1998. Greenhouse gas fluxes following tillage and wetting in a wheat-fallow cropping system. *J. Environ. Qual.* 27, 1105–1116.
- Mosier, A.R., Hutchinson, G.L., 1981. Nitrous oxide emissions from cropped fields. *J. Environ. Qual.* 10, 169–173.
- Mosier, A.R., Mack, L., 1980. Gas chromatographic system for precise, rapid analysis of nitrous oxide. *Soil Sci. Soc. Am. J.* 44, 1121–1123.
- Mosier, A.R., Duxbury, J.M., Freney, J.R., Heinemeyer, O., Minami, K., 1997. Nitrous oxide emissions from agricultural fields: assessment, measurement and irrigation. *Plant Soil* 181, 95–118.
- Ruz-Jerez, B.E., White, R.E., Ball, P.R., 1994. Long-term measurement of denitrification in three contrasting pastures grazed by sheep. *Soil Biol. Biochem.* 26, 29–39.
- Saggarr, S., Yeates, G.W., Shepherd, T.G., 2001. Cultivation effects on soil biological properties, microfauna and organic matter dynamics in Eutric Gleysol and Gleyic Luvisol soils in New Zealand. *Soil Till. Res.* 58, 55–68.
- Shepherd, T.G., 1992. Sustainable soil-crop management and its economic implications from grain growers. In: Henriques, P.R. (Ed.), *Sustainable Land Management. Proceedings of the International Conference on Sustainable Land Management*, Napier, New Zealand, November 17–23, 1991, pp. 141–152.
- Shepherd, T.G., Saggarr, S., Newman, R.H., Ross, C.W., Dando, J.L., 2001. Tillage induced changes in soil structure and soil organic matter fractions. *Aust. J. Soil Res.* 39, 465–489.
- Sitaula, B.K., Bakken, L.R., 1993. Nitrous oxide release from spruce forest soil: relationships with nitrification, methane uptake, temperature, moisture and fertilisation. *Soil Biol. Biochem.* 25, 1415–1421.
- Webb, T.H., Jessen, M.R., McLeod, M., McIntosh, P.D., Wilde, R.H., 1997. Identifying land with high class soils for protection under the RMA. *NZ Soil News* 45, 48–51.
- Williams, D.L., Ineson, P., Coward, P.A., 1999. Temporal variations in nitrous oxide fluxes from urine-affected grassland. *Soil Biol. Biochem.* 31, 779–788.